

PhD Open Days



An Integrated Approach to the Production of Extracellular Vesicles from Human Induced Pluripotent Stem Cells in Stirred-Tank Bioreactors

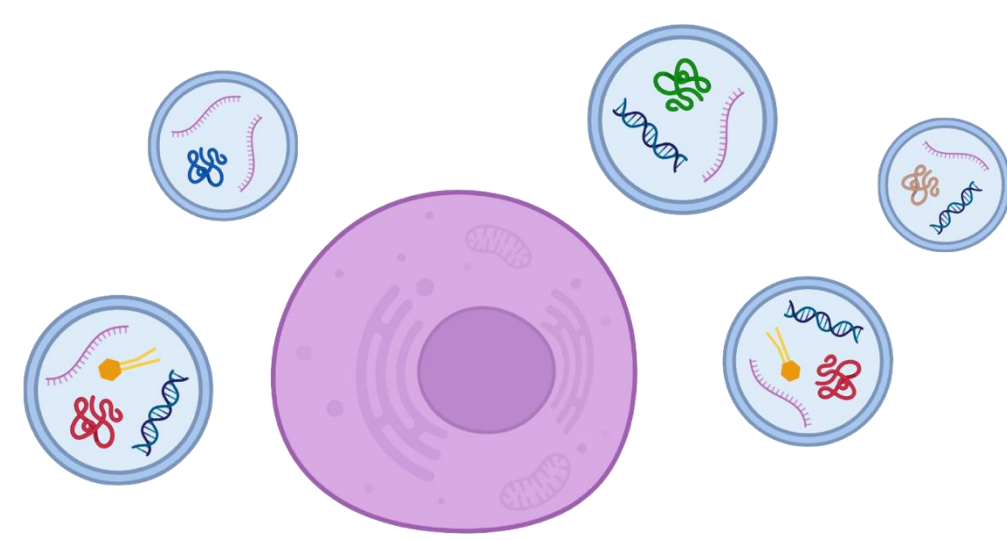
PhD Program in Bioengineering

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Relevance

Extracellular vesicles (EVs) are lipidic nanostructures secreted by cells that carry complex biological cargo composed of proteins, lipids, RNA and DNA. Depending on their cargo, **EVs** can:

- Stimulate cell proliferation
- Prevent cell death
- Promote vascularization



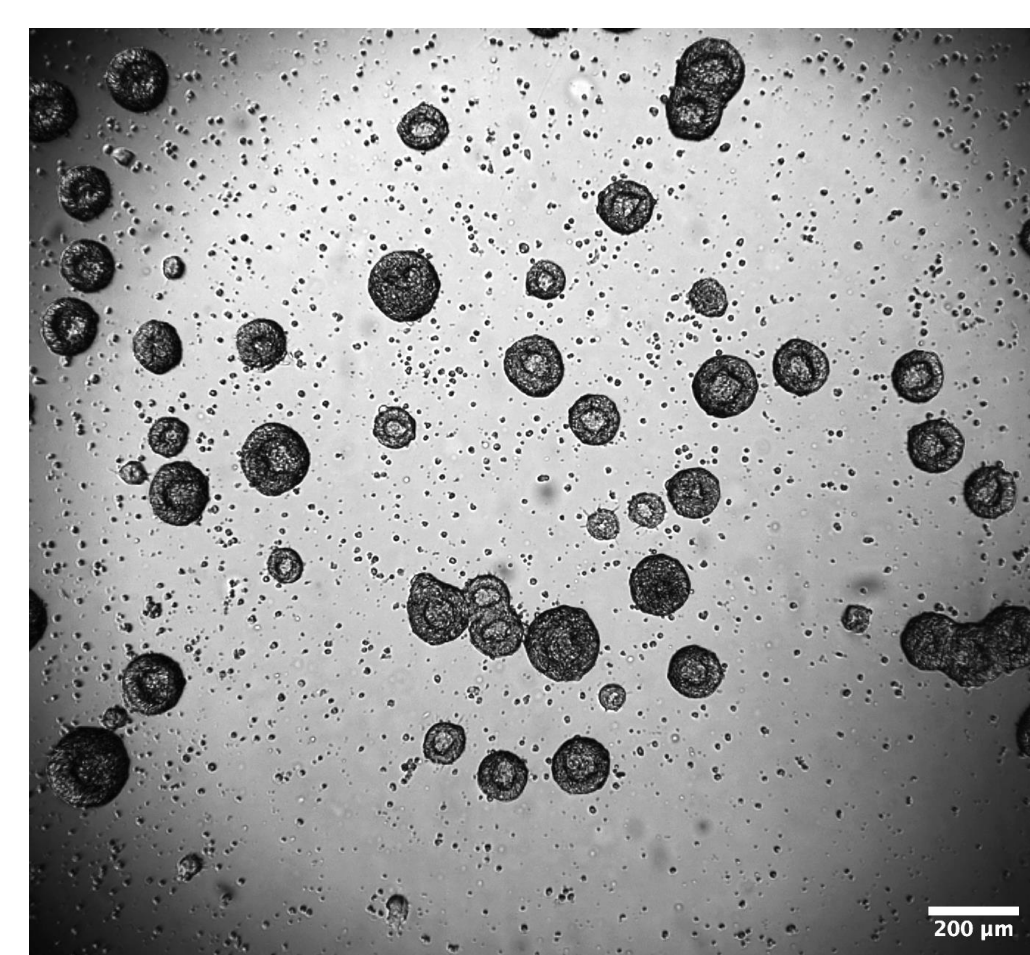
Coupled with the fact that **EVs** are not directly targeted by the immune system and can easily cross biological barriers, they have increasingly come to be seen as promising therapeutic agents for **regenerative medicine**. For example, **EVs** can be used to treat acute myocardial infarction and retinal degeneration.

EVs from **human induced pluripotent stem cells (hiPSCs)** have been observed to have higher regenerative potential than committed cell types.

Methodology

hiPSC Expansion

- Free-floating aggregates in Eppendorf's DASbox Mini Bioreactor System
- Optimization of agitation rate, medium exchange strategy and culture media (**mTeSR1**, **E8** and two formulations from **Defined Bioscience: HiDef-B8** and **A8**)
- Assessment of hiPSC pluripotency (flow cytometry and immunostaining)

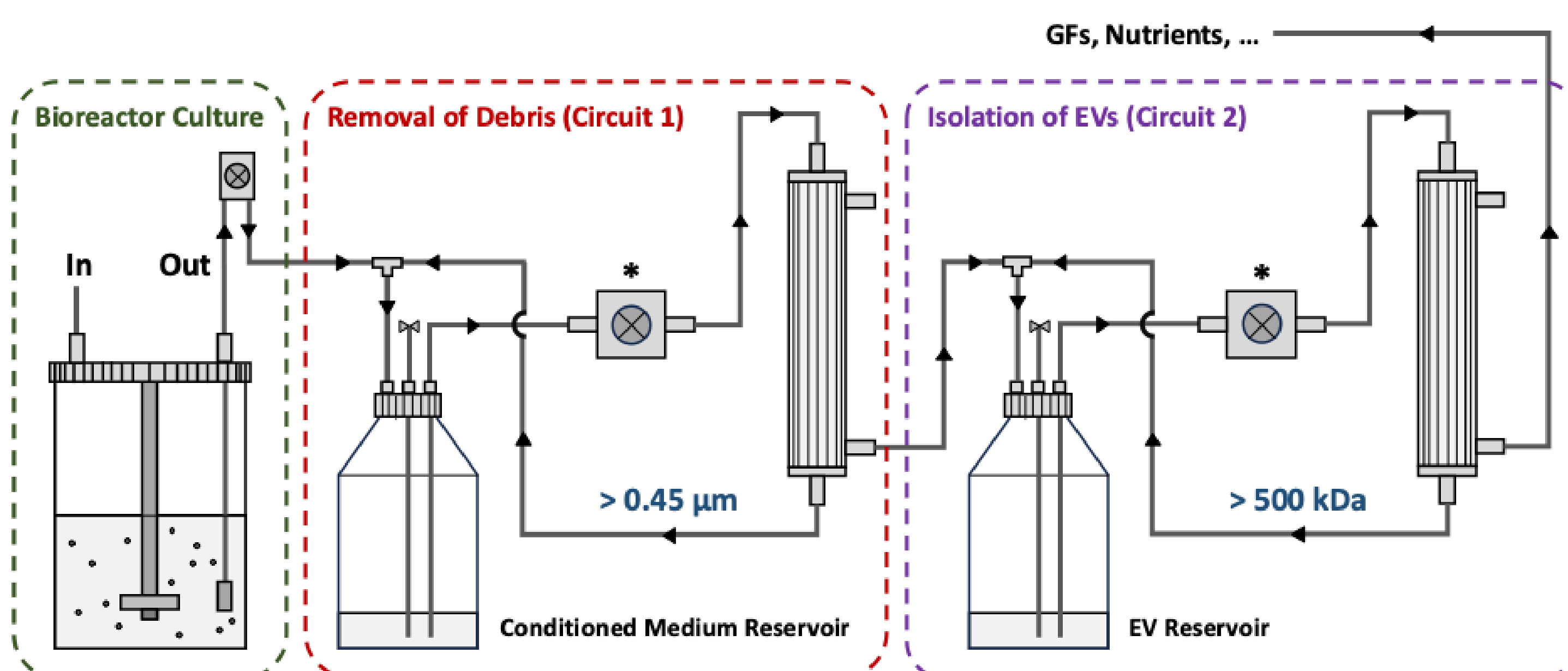


hiPSC free-floating aggregates

Two-Step Hollow Fiber TFF System

1st Pore size of 0.45 μm
Removes cell debris

2nd Pore size of 500 kDa
Retains and concentrates EVs
Removes small biomolecules



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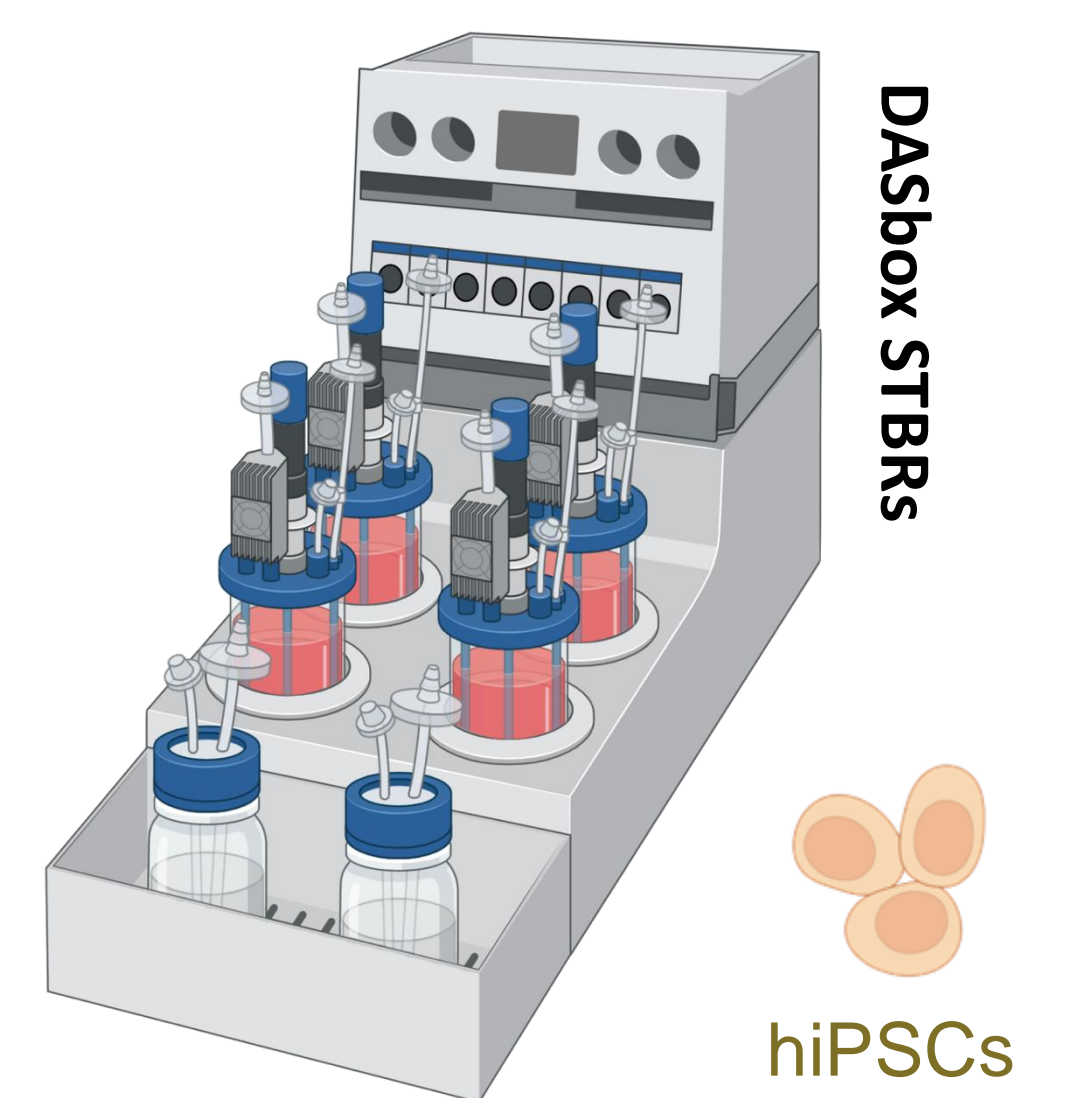
Problem

Treating a single patient can require up to 100 billion **EVs**, an amount difficult to acquire with traditional production methods. **Stirred-tank bioreactors (STBRs)** are ideal platforms for scaling up the culture of **hiPSCs** and **EV** production because they:

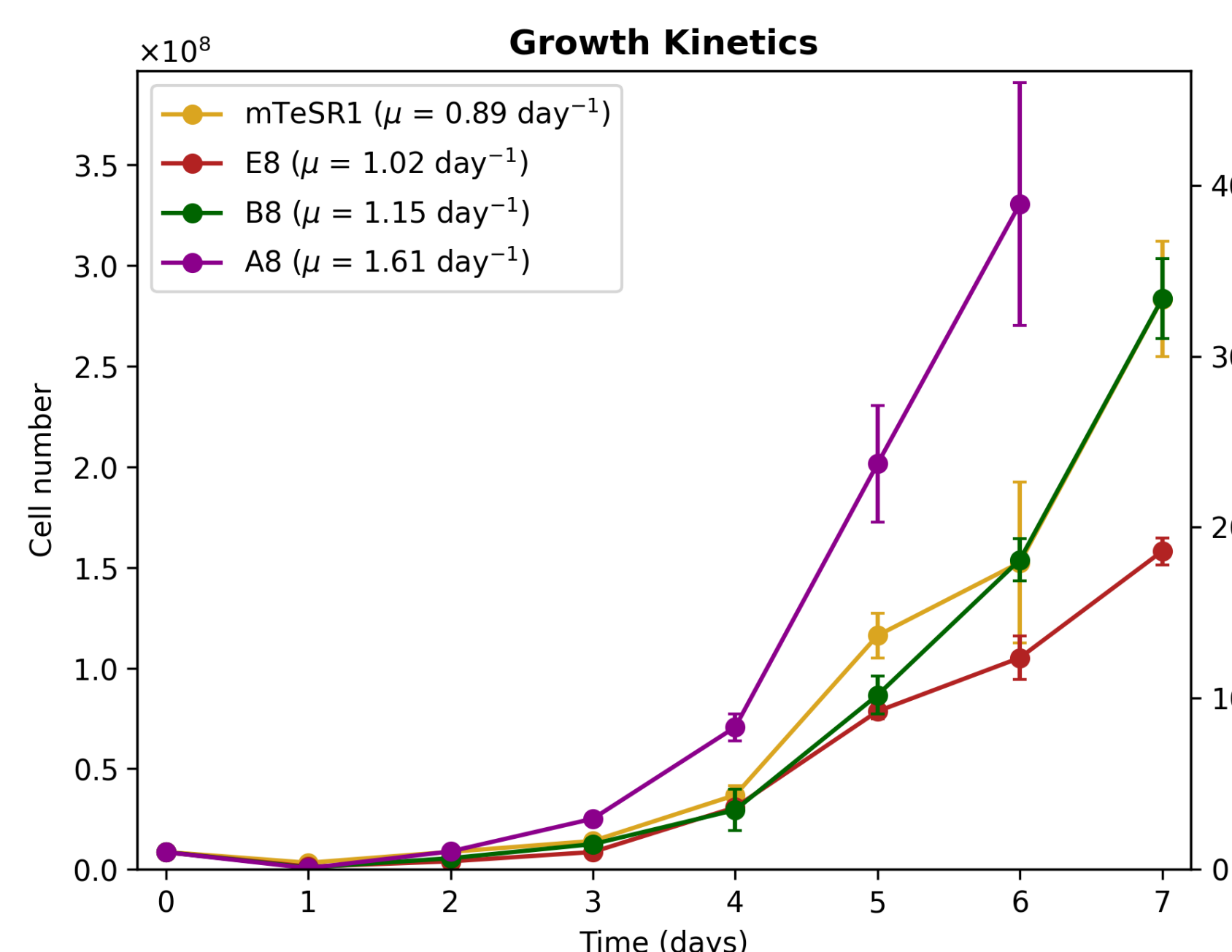
- Integrate monitoring and automation
- Allow for high cell densities
- Are highly scalable

Once **EVs** are secreted by cells they need to be recovered and purified. Among the several strategies available to achieve this, **tangential flow filtration (TFF)** is highly scalable and offers an acceptable tradeoff between yield and purity.

The **objective** of this work is to establish an **hiPSC-EV** production platform that integrates an **STBR** and a **TFF** system for **EV in-situ recovery**.



Results and Conclusions



Fold Expansion

- **mTeSR1/B8** – 33×, 7 days
- **E8** – 18×, 7 days
- **A8** – 39×, 6 days

Pluripotency Maintenance

- **mTeSR1/A8** – Expression of all markers higher than 95%
- **E8/B8** – Notably decreased expression of TRA-1-60

A8 presented improved cell survival (38%) and faster growth rate, aside from being the second cheapest medium after **B8**

Medium	Particles/mL	Total Particles
E8	$(0.82 \pm 0.13) \times 10^{10}$	$(0.68 \pm 0.11) \times 10^{12}$
A8	$(7.98 \pm 0.44) \times 10^{10}$	$(6.38 \pm 0.35) \times 10^{12}$

Particles in Medium (measured by nanoparticle tracking analysis)

- **A8** – 9.4× more particles than **E8**
- **mTeSR1** – masking effect due to large number of particles in medium

A8 presents itself as the best culture medium for optimized **hiPSC-EV** production. In the future, further analysis of particles to guarantee they are EVs and characterize their contents will be performed.